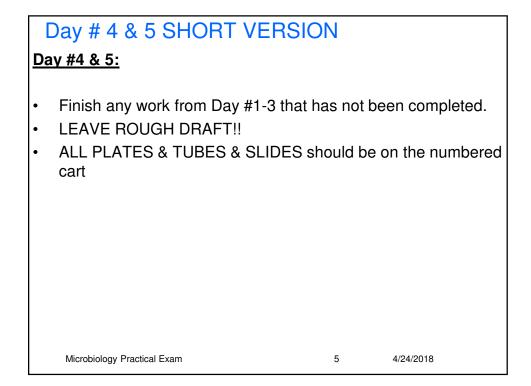
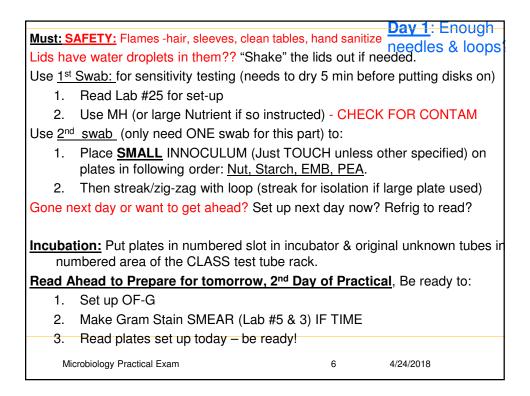


Day #3 SHORT VERSION			
<u>Day #3:</u>			
<ol> <li>Read ALL plates &amp; tubes that were set up on Day #1 or 2. If not enough time, refrigerate.</li> </ol>			
Record observations & interpretations.			
Compare plate growth to each other: Inhibited/Selective?     Differential? (Lab #12)			
Record colony morphology on nutrient (Lab #0)			
Read OF-G. (Lab #13) (& will be given TSI picture)			
2. Catalase test with H2O2 (Lab #19)			
3. If time, stain gram stain slide (Lab #5)			
<ul> <li>SHOW ME AN APPROPRIATE AREA of the slide - not too thick or thin to see shape &amp; arrangement</li> </ul>			
4. LEAVE ROUGH DRAFT!!			
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1.	Set up the following: Gone yesterday? Work at dif table.
	A. OF-G & TO (From plate w/best growth; Nut? MH –but not near disks? Day 2
2.	Make gram stain-gram stain Use FROSTED slide & pencil.
	<ul> <li>**WAX circle goes on UNDERSIDE. Leave overnight on slide warmer.</li> </ul>
3.	Read <u>Antibiotic Disk Plate</u> – Read zone sizes. If "2" zones, comment and then measure inner complete inhibition. Plate goes onto cart in "numbered" spot.
4.	Special Considerations:
	A. If contaminated/scant growth: ask me. Incubated upside down? Careful.
	B. Need to redo? Ask 1 <sup>st</sup> .
	C. Date & record observations-comment on unusual observations (ie-2 zones
	inhibition, yellow contaminant not on streak, relatively tiny growth-break
	through??) IF think CONTAMINATED H2O, oil, stick, pipet – tell me.
	D. Need more needles, wax pencils? Other supplies on table.
5.	Starch – Iodine in beaker. Don't dump I2 into sink – ONLY plate to put into "Bag".
6.	Read other plates (PEA, EMB, Nut) that were set up yesterday – NOTE Time Limits!
	A. Record OBSERVATIONS (relative ease of growth -/+/++/etc, color, - to see if
	any plates appear to be selecting and inhibiting. Selective?? Differential??)
	B. Add <u>BASIC</u> interpretations (GN, A/A, LF, oxidizer, etc. as appropriate)
	C. NOTE: Time frame for reading given – reinc Nut & EMB. Put PEA in cart spot.
	D. ***Mechanisms not needed on rough draft-but could start Final Draft & be done
L	ahead of reading.
7.	Place rough draft of lab report in report bin-have date for set up, 24 hour, etc.
8.	Put plates that are done incubating on numbered cart – even if haven't set up gram
	stain yet. <b>DON'T toss them!!</b> (Put tubes that are done in the numbered rack in the hood.)

 Prepare for tomorrow: Suggest reading info for reading tests & making smears Microbiology Practical Exam 7 4/24/2018

η.	Highlights on rough draft-meaning. Missing rough drafts? Day 3, 4, 5		
2.	Early out? Short day? Record observations (SEE procedures) & leave interpretation till tomorrow if needed.		
З.	CONTAM pipet, stick, oil? Tell me.		
4.	Special considerations:		
	A. Date & record observations-comment on unusual observations (ie-2 zones inhibition, yellow contaminant not on streak, relatively tiny growth-break through??)		
5.	Read tests: ALL plates that have incubated 48 hours should be on cart end of period.		
	A. Finish Nutrient – form, margin, elevation, size in mm, color. Put in hood.		
	B. Finish EMB – compare ease of growth & size to Nutrient. Color? Basic interp? Put in hood.		
	C. Finish Starch – Add iodine, read, place plate in hood, NOT cart so doesn't spill.		
	D. OF- G: Compare to <u>unused tubes</u> . Color & basic interp. Put tubes in rack on cart.		
6.	Catalase:H2O2 in old petri lid. Use stick; in lysol beaker when done. Observe & Interp.		
7.	Slides – Stain if time today. (Should have made smears yesterday)		
	A. Slide warmers. Frosted ends-PENCIL. Put "sliders" in disinfectant beaker. Gram iodine in <u>dropper bottle</u> (not beaker). Leave iodine on table, NOT in bin.		
	B. Redo until satisfied		
	C. <u>SHOW ME APPROPRIATE AREA OF SLIDE</u> , – not too thick/thin to tell shape & arrange, in focus, when READY put name on "WAITING LIST"		
	D. When done or waiting to read gram stain, place slides on top of your pile of "done" plates.		
8.	Rough Draft – LEAVE in bin. Use phone to take pic of rough draft if you want a copy.		
9.	Final Report Specific & detailed interpretations (based on previously recorded observations) & mechanisms – HOW/WHY/CAUSE of ALL TYPES results		
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